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(54) Title: CARBOXYAMIDE DERIVATIVES AS SELECTIVE INHIBITORS OF PATHOGENS

(57) Abstract: The present invention relates to the use of carboxamide compounds as selective inhibitors of pathogens, particularly viruses and, more particularly, herpesviridae. Surprisingly, these compounds show reduced side effects in comparison with previous antiviral compounds. Thus, a novel method for preventing or treating infections by pathogens, particularly herpesviridae is provided.

Carboxamide derivatives as selective inhibitors of pathogens**Description**

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The present invention relates to the use of carboxamide compounds as selective inhibitors of pathogens, particularly viruses and, more particularly, herpesviridae. Surprisingly, these compounds show reduced side effects in comparison with previous antiviral compounds. Thus, a novel method for preventing or treating infections by pathogens, particularly herpesviridae is provided.

Human Cytomegalovirus (HCMV) is a highly species specific β -herpesvirus. Primary infection of healthy children and adults is usually asymptomatic, with a minority of cases developing a mononucleosis-like syndrome. In contrast, congenital infection (U.S 0.2%-2.2% per live birth; approx. 40,000 per year) leads to several neurological defects in 10 to 15% of infected neonates. Immunocompromised patients are another group of hosts facing serious diseases caused by HCMV infection or reactivation of a persistent infection. Up to 40% of AIDS patients, for example, develop retinitis, pneumonitis, gastroenteritis or disseminated HMCV disease. Allograft recipients (20,000 allograft transplantations per year in US) are often infected (or superinfected) by virus from the transplanted organ. Clinical symptoms in the postransplant period include prolonged fever, leukopenia, thrombocytopenia, atypical lymphocytosis, elevated hepatic transaminases and decreased graft survival. In bone marrow transplantations, HCMV infection has been associated with high mortality rates (80-90% for untreated HCMV pneumonia), which have been reduced by newer antiviral agents to 10-20% (reviewed in Britt J.B. and Alford C.A., 1996, *Cytomegaloviruses*, pp.2494-2523. In B.N.Fields, D.M.Knipe, P.M.Howley (ed.) *Fields Virology*, Lippencott-Raven Pub., Philadelphia).

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Ganciclovir is available for intravenous and oral administration and as an implant in the case of retinitis. Toxicities include leukopenia and thrombocytopenia. Foscarnet (phosphonoformic acid) exhibits considerable renal toxicity and is only available in intravenous form, which is also true for Cidofovir. Although treatment of HCMV-induced disease has been substantially improved with these inhibitors of the viral DNA polymerase and preemptive or early antiviral therapy in transplant patients (Hebart, H. et al., Drugs, 1998, 55:59-72), there is room for improvement in the toxicity profile. Especially in the treatment of retinitis in AIDS patients, where CMV infection has to be controlled for long periods of time and replication will resume once Ganciclovir and Foscarnet are removed, new classes of drugs are needed with better oral bioavailability and activity against emerging Ganciclovir-and Foscarnet-resistant strains.

Leflunomide, an immunomodulatory drug used in rheumatoid arthritis, was previously found to inhibit HCMV replication in cell culture. The antiinflammatory and immunosuppressive properties of Leflunomide have been demonstrated in animal models of autoimmune disease and organ transplant rejection (Reviewed in J.Rheumatology (1998) 25:20, Agents Actions (1991) 32:10) and it was recently approved for use in rheumatoid arthritis in the US (Scrip (1998) (2370):22). The proposed basis for the antiproliferative action of Leflunomide, for example, in mitogen-stimulated human T-lymphocytes (JBC (1998) 273:21682), is the suppression of the de novo pathway of pyrimidine synthesis. The active metabolite of Leflunomide is a noncompetitive inhibitor of Dihydroorotate dehydrogenase (DHODH) (Biochem. Pharmacol. (1995) 50: 861; J.Pharmacol.Exper.ther.(1995) 275:1043; JBC (1995) 270:22467, Eur.J.Biochem.(1996) 240:292; Biochemistry (1996) 35:1270), a mitochondrial enzyme which catalyzes the rate-limiting step in this biosynthetic pathway. Most research groups favor the inhibition of DHODH as the major mechanism for Leflunomide-induced cytostatic effects (cited in JBC 273, 21682), however, inhibition of tyrosine kinases by Leflunomide

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has been demonstrated in animal models and proposed to be a second important mode of action in immunosuppression (J.Immunol.(1997) 159:22; JBC (1995) 270:12398; Biochem.Pharmacol.(1996) 52:527, J.Immunol.159:167). Tyrosine kinase inhibition is not affected by the
5 addition of uridine, while DHODH inhibition can be reversed by uridine. In the case of Brequinar Sodium, which is another inhibitor of DHODH and an antimetabolite tested for cancer therapy (Cancer Res.(1986) 46:5014, Cancer Chemother.Pharmacol.(1993) 32:64), the finding that only antiproliferative effectes were reversed by uridine led to a patent claiming
10 the reduction of non-immunosuppressive side effects by coadministration of uridine (Williams J.W., Chong, A., Xu, X.: WO 98/13047 (1994)).

Surprisingly, it was found that carboxamide derivatives can selectively inhibit kinases from pathogens, particularly from herpesviridae such as
15 HCMV, without significantly inhibiting DHODH. Particularly, these compounds show a pronounced reduction in HCMV replication but do not inhibit purified recombinant human DHODH in an in vitro assay. This surprising uncoupling of antipathogen efficacy and inhibition of the de novo pyrimidine synthesis suggests that the subject carboxamide derivatives are
20 free of the side effects associated with the antiproliferative and immunosuppressive properties of prior art medicaments such as Leflunomide.

These compounds have been shown previously to inhibit platelet-derived
25 growth factor receptor activity (WO 95/19169 Sugen, Inc.), to have antibacterial activity (US 3,303,201) and to inhibit HIV reverse transcriptase (WO 96/16675 Rega Institut). A selective activity of these compounds against kinases from pathogens such as bacteria, protozoa or viruses, particularly without undesired side effects, has not yet been described.

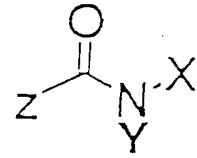
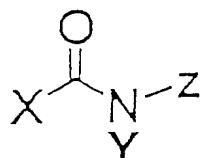
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Thus, the present invention refers to the use of compounds of the general formulae (Ia), (Ib), (Ic) or (Id):

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(Ia)

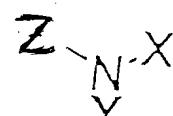
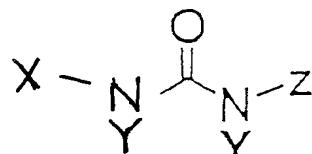
(Ib)



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(Ic)

(Id)



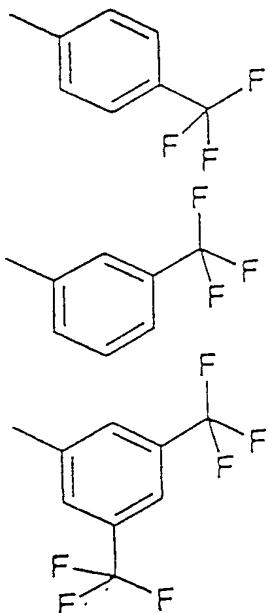
wherein X and Z are substituents comprising an aromatic or heteroaromatic ring system for the manufacture of an agent against infectious diseases. For example, X and/or Z may be an aromatic radical, preferably a phenyl radical which is unsubstituted or which carries at least one substituent, e.g. 1-4 substituents which may be selected from hydroxy, cyano, nitro, halo, e.g. F, Cl, Br, I, C₁-C₄ alkyl, C₁-C₄ haloalkyl, C₁-C₄ alkoxy, C₁-C₄ haloalkoxy, C₁-C₄ alkylthio, C₁-C₄ haloalkylthio, carboxy, carboxy-C₁-C₄-alkyl, carboxy-aryl (or heteroaryl), amino carbonyl-C₁-C₄ alkyl, amino carbonyl - aryl (or heteroaryl) and aryl (heteroaryl), Y is hydrogen or C₁-C₄ alkyl.

Especially preferred are compounds, wherein X and/or Z is an aromatic or heteroaromatic radical, e.g. a phenyl radical having at least one, particularly one, two or three C₁-C₄ haloalkyl substituents, e.g. C₁ haloalkyl substituents such as -CF₃, -CHF₂ and -CH₂F.

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For example, X and/or Z may be selected from radicals represented by formulae (IIa-c):

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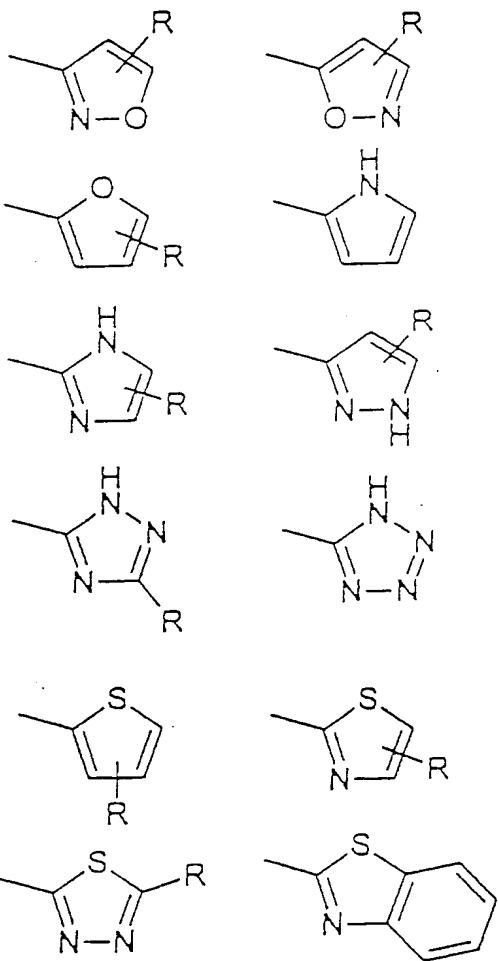
20 Y is preferably hydrogen or C₁-C₂ alkyl, more preferably hydrogen.

In a further preferred embodiment, X and/or Z comprises a heterocyclic ring which may contain one or several heteroatoms such as oxygen, nitrogen and/or sulfur, preferably a 5-membered heterocyclic ring which may be selected from pyrrole, pyrazole, imidazole, 1,2,3-triazole, tetrazole, oxazole, isoxazole, thiazole, isothiazole, 1,2,4-thiadiazole, 1,3,4-thiadiazole, thiophene, furan, indole and 3-thiaindole, a 6-membered heterocyclic ring which may be selected from pyridine, pyran, pyrimidine, pyridazine, pyrimidine and pyrazine, or a bi- or polycyclic heteroaromatic ring such as indazole, imidazole, chinoline or isoquinoline. The heterocyclic ring can be mono, di, tri or tetrasubstituted with substituents as defined above.

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For example, X and/or Z may be selected from radicals represented by formulae (IIIa-k):

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wherein R is at least one substituent, e.g. one or two substituents selected from halo, C₁-C₃ alkyl, C₁-C₃ alkoxy or aryl, e.g. phenyl.

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The aromatic and/or heteroaromatic ring of X and Z may be directly linked to the central structural element of the compounds (Ia-d), i.e. by a covalent

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bond. Alternatively, the linkage may comprise an alkylene-group, preferably a C₁-C₄ alkylene group, e.g. a C₁ or C₂ alkylene group.

Surprisingly, compounds (Ia), (Ib), (Ic) and (Id) have a selectivity for kinases from pathogens compared with the cellular enzyme DHODH. Thus, the compounds are suitable for the preparation of an agent against infectious diseases, particularly viral infections, more particularly, against infections by herpesviruses. The herpesviruses may be selected from human herpesviruses and herpesviruses from other mammals, such as bovine, equine, porcine and pongine herpesviruses. Suitable herpesviruses are selected from α -herpesviruses, e.g. simplexviruses such as herpes simplex virus 1, herpes simplex virus 2, bovine herpesvirus 2, cercopithecine herpesvirus 1 or varicellaviruses such as varicella zoster virus, porcine herpesvirus 1 (pseudorabiesvirus) bovine herpesvirus 1 and equine herpesvirus 1 (equine abortion virus). Further, the herpesvirus may be selected from β -herpesviruses, e.g. cytomegaloviruses such as human cytomegalovirus and from roseoloviruses, such as human herpesvirus 6, human herpesvirus 7 or aotine herpesviruses 1 and 3. Further, the herpesviruses may be selected from γ -herpesviruses, e.g. from lymphocryptoviruses such as Epstein-Barr virus, cercopithecine herpesvirus 2 or porcine herpesvirus 1, or from rhadinoviruses such as human herpesvirus 8, ateline herpesvirus 2 or saimiriine herpesvirus 1, or preferably, the virus is selected from human herpesvirus 1 (HSV-1), varicella zoster virus (VZV) or human cytomegalovirus (HCMV).

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Further, it was surprisingly found that the compounds are active against Foscarnet-resistant HCMV strains. Thus, they are likely to have a mode of action which is different from the HCMV drugs on the market, making them valuable tools for combination therapy approaches, e.g. for combination therapies together with viral DNA polymerase inhibitors. Furthermore, it was found that the compounds are potent inhibitors of ganciclovir-resistant virus strains, particularly ganciclovir-resistant HCMV strains.

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The compounds of the formula I are suitable for the manufacture of an agent for the prophylaxis and/or treatment of a viral infection. This prophylaxis or treatment comprises administering a pharmaceutical composition containing as an active agent a pharmaceutically effective amount of a compound of the general formula I to a subject, preferably a human, in need thereof, e.g. a subject suffering from a herpesvirus infection or a subject which is in need of a prophylactic administration to avoid the outbreak of a herpesvirus infection. The pharmaceutical composition may contain suitable diluents, carriers and auxiliary agents. Further, the composition may also contain other pharmaceutically active agents, e.g. antiviral agents. The pharmaceutical composition may be suitable for oral, parenteral, e.g. intradermal, intravenous or intramuscular, rectal, nasal and topical applications. The composition may be an injectable solution, ointment, cream or spray. Further, the composition may have retardation properties, i.e. showing a delayed release of the active agent.

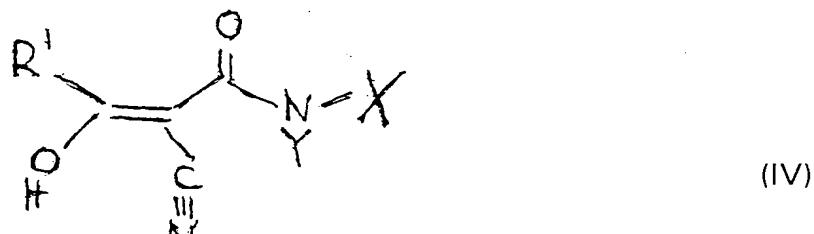
The dosage of the active agent depends on the specific compound being administered, the type and the severity of the viral infection. For example, a dosage from 0.01 mg to 100 mg per day and per kg/body weight for the active agent is suitable.

The efficacy of the compounds of formulae (Ia), (Ib), (Ic) and (Id) against enzymes from pathogens, e.g. viral kinases, may be determined in an in vitro enzyme inhibition test. On the other hand, the anti-viral effect can also be determined directly in a cell culture assay. A preferred and new cell culture assay is described in Example 2.2 and comprises the use of cells infected with a recombinant human cytomegalovirus carrying a reporter gene, e.g. the GFP-gene. The reporter gene is preferably inserted into the viral genome in a manner that viral replication is still possible. For example, the reporter gene may be inserted into the HCMV gene region encoding the open reading frames US9 and US10.

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A further embodiment of the present invention refers to the use of compounds of the general formula (IV):

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wherein X and Y are defined as for the compounds (Ia-d) and R' is selected from C₁-C₄ alkyl and C₃-C₄ cycloalkyl, particularly CH₃, for the preparation of an agent against ganciclovir-resistant virus strains. The compounds may be present in the form of a physiologically acceptable salt, e.g. an alkali metal, ammonium or substituted ammonium salt, particularly the sodium salt or the salt of a basic amino acid such as lysine. The compounds of formula (IV) may be classified as leflunomides. Surprisingly, these leflunomides are potent inhibitors of ganciclovir-resistant virus strains, particularly ganciclovir-resistant herpes virus strains and more particularly ganciclovir-resistant HCMV strains.

The compounds of the formula (IV) are suitable for the manufacture of an agent for the prophylaxis and/or treatment of a viral infection. This prophylaxis or treatment may be carried out as described above for compounds of the formulae (Ia) and (Ib).

Furthermore, the invention is to be explained by the following figures and examples.

30 Fig. 1 is a list of compounds of the present invention having anti-viral activity.

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Fig. 2 is a diagram showing the efficacy of compounds of the present invention against a ganciclovir-resistant virus strain.

Example 1

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Synthesis of substituted anilides of heterocyclic carboxylic acids

A general procedure is given here suitable for acylation of substituted anilines with heterocyclic carboxylic acids. The aromatic rings can be mono, di, tri, or tetrasubstituted with e.g. alkyl, substituted alkyl, aryl, hydroxy, halogen, nitro, carboxy, thiomethyl groups. The heterocyclic ring can be mono, di, tri, or tetrasubstituted with e.g. alkyl, substituted alkyl, aryl, hydroxy, oxo, halogen, nitro, thiomethyl groups.

15 General procedure

1-10 mmol aniline was dissolved in dry 10-30 mol of dry pyridine and the solution was cooled to -20°C. 0.5 eq. of phosphorous trichloride was dropped in with vigorous stirring while the temperature was kept under -15°C. After 10-15 min the carboxylic acid was added slowly with stirring. The reaction mixture was allowed to warm to room temperature and stirred overnight, then warmed to 40°C until 30 min. Pyridine was removed in vacuum and the residue was taken up in 1N hydrochloric acid and ethyl acetate. The organic phase was washed with 5% hydrocarbonate solution and distilled water. After drying over sodium sulfate the solvent was evaporated and the residue purified via filtrating through silica layer, or chromatographed or recrystallized, if necessary.

All reagents and intermediates were purchased from Sigma-Aldrich.

30

For example, the following compounds were obtained:

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(1) Thiophene-2-carboxylic acid (3-trifluoromethyl phenyl)amide M.p.:
151-152°C R_f:0.73 (Ethylacetate:Hexane = 1:1)

5 (2) Furane-2-carboxylic acid (3-trifluoromethyl phenyl)amide
M.p.: 108-109°C R_f:0.73 (Ethylacetate:Hexane = 1:1)

(3) Thiophene-2-carboxylic acid (3,5 bis-(trifluoromethyl) phenyl)amide
M.p.: 148°C R_f:0.8 (Ethylacetate:Hexane = 1:1)

10 (4) Furane-2-carboxylic acid (4-trifluoromethyl phenyl)amide
M.p.: 108-109°C R_f:0.73 (Ethylacetate:Hexane)

(5) Furane-2-carboxylic acid (3,5 bis-(trifluoromethyl) phenyl)amide
M.p.: 124-125°C R_f:0.85 (Ethylacetate)

15 (6) 2-Cyano-3-hydroxy-crotonic acid (4-trifluoromethyl phenyl) amide .

According to the general procedure as described above, further compounds
of the formula I can be prepared, e.g. by using corresponding pyrrol,
20 pyrrolidine or oxo-pyrrolidine carboxylic acids as starting materials. A list of
such compounds is shown in Fig. 1.

Example 2

25 **Anti-viral activity of compounds according to formulae (Ia) and (Ib)**

2.1 DHODH assay

The cDNA for human DHODH (accession number M94065) was obtained
via PCR with EST clone AA173225 (from RZPD in Berlin) and
30 oligonucleotides 5'-CTG AAT TCA AAT TAC CGT GGA GAC ACC TGC
AAA AGC GGG CCC AG-3' (this oligo provides the coding sequence for the

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first 5 amino acids missing in the EST clone) and 5'-AGC TCG AGT CAC CTC CGA TGA TCT GCT CCA AT-3'. The PCR product was subcloned with EcoRI and Xhol into vector pGEX 5x1 (Pharmacia, Gene (1988) 67:31) for expression as a GST-fusion protein in bacteria. Recombinant GST-DHODH was purified on glutathione-sepharose (method described in Gene (1988) 67:31) and dialyzed against 20 mM Tris pH 7.5, 0.1% Triton X100. The assay was basically performed as described by Copeland et al. ("NBT assay", Arch.Biochem.Biophys.323:79, 1995) with slight modifications: 20 µl of a 5x ubiquinone mixture (1 mM Ubiquinone Q10, 0.5% Triton X100, 10 500 mM Tris pH 7.5), 2 µl of 10 mM NBT (nitroblue tetrazolium), 2-10 µl of DHODH-GST (depending on the batch of 1-liter bacterial preparation), 91- 83 µl of water and 5 µl of compound or DMSO were added to each well of a 96-well microtiter plate. After mixing and incubation of the reactions for 15 5 min at 37°C, 5 µl of 10 mM L-dihydroorotate (or 5 µl of water in case of the background controls) were added to start the reactions. After incubation at 37°C for 60 to 90 min, ODs were read at 595 nm with a Biorad microplate reader. Readings were in duplicate, controls without L-dihydroorotate were performed as single measurements. Activity was normalized with values from DMSO controls.

20

2.2 Virus Replication Assay

Cell culture and virus

Primary human foreskin fibroblasts (HFF) were cultivated in MEM containing 25 5% (v/v) fetal calf serum. Infection analysis was restricted to cell passage numbers below twenty. Human cytomegalovirus strain AD169 (ATCC) was grown in HFF cells and quantitated for infectivity by the plaque reduction assay. Aliquots were stored at -80°C.

30 Construction of recombinant cytomegalovirus

For construction of a recombination vector, two linker sequences were inserted into the pBlueScribe vector pBS + (Stratagene): the first contained

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restriction sites for NheI, SphI, PstI and BglII followed by a loxP sequence (ATAACTTCGTATAGCATACATTATACGAAGTTAT) and was introduced into PstI/XbaI sites of the vector; the second contained another loxP sequence followed by restriction sites HpaI, ClaI and PmeI and was introduced into BamHI/Asp718 sites. A gene cassette comprising of a "humanized" version of the ORF coding for GFP (gfp-h) under the control of the HCMV enhancer/promoter and the Ptk/PY441 enhancer-driven neoR selection marker was excised from plasmid pUF5 (Zolotukhin et al., 1996, J.Virol.70, 4646-4654) and inserted into the recombination vector via BglII sites.

At the 5' and 3'-positions of this loxP-flanked gene cassette, two HCMV sequences with homology to the gene region containing the open reading frames US9 and US10 were inserted. For this, viral sequences were amplified from template pCM49 (Fleckenstein et al., 1982, Gene 18, 39-46) via PCR in a 35-cycle program (denaturation 45 sec at 95 °C, annealing 45 sec at 55 °C and elongation 2 min at 72 °C) by the use of Vent DNA polymerase (New England Biolabs). A US10-specific sequence of 1983 bp in length was generated using primers US10[200900]SphI (GCTCACTAGTGGCCTAGCCTGGCTCATGGCC) and US10[198918]PacI (GTCCTTAATTAAGACGTGGTTGTGGTCACCGAA) and inserted at the vector 5' cloning position via SphI/PacI restriction sites (see bold-print). A US9-specific sequence of 2010 bp was generated using primers US9-3'PmeI (CTCGGTTAACGACGTGAGGCGCTCCGTCACC) and US-5' ClaI (TTGCATCGATACGGTGTGAGATACCACGATG) inserted at the vector 3' cloning position via PmeI/ClaI restriction sites.

The resulting construct pHM673 was linearized by the use of restriction enzyme NheI and transfected into HEK cells via the electroporation method using a Gene Pulser (Bio-rad; 280 V, 960 µF, 400 Ω). After 24 h of cultivation, cells were used for infection with 1 PFU/ml of HCMV strain AD169. Selection with 200 µg/ml G418 was started 24 h post infection.

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Following 3 weeks of passage in the presence of G418, GFP fluorescence could be detected in most of the infected cells. Plaque assays were performed with infectious culture supernatant on HFF cells and single virus plaques were grown by transfer to fresh HFF cells cultured in 48-well plates. DNA was isolated from cells of 32 fluorescence-positive wells and confirmed for the presence of recombinant virus by PCR. For this, primers US9[198789] (TGACGCGAGTATTACGTGTC) and US10[199100] (CTCCTCCTGATATGCGGTT) were used resulting in an amplification product of 312 bp for wild-type AD169 virus and approximately 3.5 kb for recombinant virus.

Plaque assay

HFF cells were cultivated in 12-well plates to 90-100% confluence and used for infection with dilutions of virus-positive cell culture supernatants. Virus inoculation was performed for 90 min at 37°C under occasional shaking before virus was removed and the cell layers were rinsed with PBS. Overlays of MEM 5% (v/v) fetal calf serum and 0.3% (w/v) agarose were added to each well and all samples were incubated at 37°C in a 5% CO₂ atmosphere for approximately 12 days. Finally, overlays were removed and the formation of foci was visualized by staining with 1% crystal violet in 20% ethanol for 1 min. After repeated rinsing with PBS, plates were air-dried at room temperature and plaque numbers were counted with a light microscope. For the recombinant AD169-GFP virus, quantification of plaque assays could also be performed without crystal violet staining by a direct counting of the amount of green fluorescent plaques using fluorescence microscopy.

Antiviral compounds

The reference compounds used for antiviral studies, ganciclovir (GCV, Cymeven), foscarnet sodium (FOS, Foscavir) and cidofovir (CDV, Vistide) were purchased from Syntex Arzneimittel (Aachen, Germany), Sigma-

- 15 -

Aldrich (Germany) and Pharmacia & Upjohn S.A. (Luxembourg), respectively. Stocks were prepared in aqueous solution and stored at -20°C. The test compounds were dissolved in DMSO and aliquots were stored at -20°C.

5

GFP infection assay

HFF cells were cultivated in 12-well plates to 90-100% confluence and used for infection with 0,5xTCID₅₀ of AD169-GFP virus. Virus inoculation was performed for 90 min at 37°C with occasional shaking before virus 10 was removed and the cell layers were rinsed with PBS. Infected cell layers were incubated with 2 ml of MEM containing 5% (v/v) fetal calf serum and optionally of the respective test substances or DMSO as control. Infected cells were incubated at 37°C in a 5% CO₂ atmosphere for 7 days and harvested by trypsinization and centrifugation. 200 µl of lysis buffer (25 mM 15 Tris pH 7.8, 2 mM DTT, 2 mM trans-1,2-diaminocyclohexane-N,N,N',N'-tetraacetic acid, 1% Triton X-100, 10% glycerol) was added to each cell pellet and lysis was achieved by incubation for 10 min at 37°C followed by a 30-min incubation at room temperature on a shaker. Lysates were 20 centrifuged for 5 min at 15.000 rpm in an Eppendorf centrifuge to remove cell debris. Supernatants were transferred to an opaque 96-well plate for automated measuring of GFP signals in a Victor 1420 Multilabel Counter (Wallac). GFP units were converted to percent inhibition values relative to DMSO controls (set at 100% GFP expression).

25 **Indirect immunofluorescence analysis**

Cells were either grown on Lab-Tek Permanox slides (Nunc) or harvested from 6-well plates, spotted onto glass slides with marked rings (Medco) and fixed by a 15-min treatment with 3% formaldehyde in PBS followed by permeabilization for 15 min in 0.1% Triton X-100 in PBS at room 30 temperature. Blocking was achieved by incubation with Cohn Fraction II/III of human gamma-globulin (Sigma; 2 mg/ml) for 30 min at 37°C. The IE1/IE2-specific primary antibody MA810 (Chemicon International, Inc. CA,

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USA; dilution 1:10.000) was incubated for 90 min, the secondary antibody (tetramethyl rhodamine [TRITC]-coupled anti-mouse antibody, Dianova, dilution 1:100) for 45 min at 37°C before analysis by fluorescence microscopy. In addition to indirect TRITC staining of IE1/IE2 proteins, GFP 5 signals could be detected directly via the fluorescence isothiocyanate (FITC) channel. Nuclear counterstaining was carried out using Vectashield mounting medium including DAPI (Vector Laboratories, Burlingame, CA).

2.3 Results

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First, the ability of the compounds to inhibit the de novo pathway of pyrimidine biosynthesis, possibly leading to antiproliferative and immunosuppressive effects *in vivo*, was tested by determining DHODH activity *in vitro* in the presence of 20 µM of the test compounds. The 15 maximum inhibition of DHODH was 26%, whereas the active metabolite of Leflunomide inhibited at least 80% of DHODH activity under test conditions. Compounds (1) and (3), two compounds with a thiophene ring, were very effective in inhibiting HCMV replication at concentrations of 20 and 100 µM, while (2) which is identical to (1), except that it contains a furane ring 20 instead of a thiophene ring, showed slightly lower reduction at 20 µM concentration. Compounds (4) and (5) inhibited HCMV significantly at the higher, but not at the lower concentration.

Table 1 shows the compounds and lists which percentage of DHODH 25 activity is inhibited at a concentration of 20 µM. Values are averages of duplicate measurements from two different experiments. Percentage of HCMV inhibition at 20 µM and 100 µM (two different experiments) of the substances are also shown. Values are averages of duplicate measurements.

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Table 1

	DHOD inhibition [%]	HCMV inhibition [%]	
Compound	20 μ M	100 μ M	20 μ M
(1)	6 \pm 10	99 \pm 0	90 \pm 4
(2)	5 \pm 4	96 \pm 1	73 \pm 10
(3)	11 \pm 9	100 \pm 0	99 \pm 0
(4)	12 \pm 9	92 \pm 1	9 \pm 26
(5)	15 \pm 11	98 \pm 0	39 \pm 12

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The compounds shown in Fig. 1 also have significant anti-viral activity in the assay system as described above.

Example 3

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Anti-viral activity against ganciclovir-resistant virus strains

3.1 Isolation of Drug-Resistant Virus

20 A series of laboratory variants of AD169-GFP virus with resistance against ganciclovir (GCV) was generated. HFF cells were infected in 12-well plates with MOI 0.002 and incubated with 1 μ M of GCV. GFP-expression in infected cells was monitored microscopically and the supernatants of positive wells were transferred to fresh cells weekly. Thereby GCV concentrations were increased stepwise (1- μ M increase in each step) up to 25 the point where the total virus replication became critical and resistant virus grew out in individual wells. Using supernatants of these wells, two rounds of plaque purifications were performed on HFF cells. Finally, GCV-resistant

- 18 -

viral clones (e.g. AD169-GFP314) were isolated which were able to replicate in the presence of 10 µM of GCV.

3.2 Virus Replication Assay

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Plaque purification and plaque reduction assay

HFF cells were cultivated in 12-well plates to 90-100% confluence and used for infection with dilutions of virus stocks (i.e. AD169-GFP or AD169-GFP314). Virus inoculation was performed for 90 minutes at 37°C under occasional shaking before virus was removed and the cell layers were rinsed with PBS. Overlays of MEM containing 5% (v/v) fetal calf serum and 0.3% (w/v) agarose were added to each well. The plates were incubated at 37°C in a 5% CO₂ atmosphere for 8-12 d. For plaque purification of GFP-expressing viruses, plates were used for fluorescence microscopy, GFP-positive plaques were picked from the overlays and transferred to fresh cells for virus multiplication. For plaque reduction assays, antiviral compounds were incubated in the overlays after infection. Overlays were removed and plaque formation was visualized by staining with 1% cristal violet in 20% ethanol for 1 min. After repeated rinsing with PBS, plates were air-dried at room temperature and plaque numbers were counted with a light microscope. For the GFP-expressing recombinant viruses, quantification of plaque reduction assays could be performed alternatively, without cristal violet staining, by a direct counting of the numbers of green fluorescent plaques using fluorescence microscopy.

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- 19 -

one of the respective test compounds. Infected cells were incubated at 37°C in a 5% CO₂ atmosphere for 7 d. For lysis, 200 µl of lysis buffer (25 mM Tris pH 7.8 2 mM DTT, 2 mM trans-1,2-diaminocyclohexane-N,N,N',N'-tetraacetic acid, 1% Triton X-100, 10% glycerol) was added to each well and incubated for 10 min at 37°C followed by a 30-min incubation at room temperature on a shaker. Lysates were centrifuged for 5 min at 15,000 rpm in an Eppendorf centrifuge to remove cell debris. 100 µl of the supernatants were transferred to an opaque 96-well plate for automated measuring of GFP signals in a Victor 1420 Multilabel Counter (Wallac).

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3.3 Results

GCV-resistant HCMV is sensitive to compound (1) or compound (6).

15 HFF cells were cultivated in 12-well plates and infected with the indicated concentrations of GFP-expressing variants of HCMV, AD169-GFP (parental) or AD169-GFP314 (GCV-resistant). Immediately after the infection, the following chemical compounds were added to the culture media: DMSO 0.07% (A), 0.14% (B), ganciclovir (GCV) 10 µM (A and B), compound (6) 20 35 µM (A) and 70 µM (B), and compound (1) 35 µM (A) and 70 µM (B). Seven days postinfection infected cells were harvested and used for GFP quantification. The results for parental virus AD 169-GFP and a GCV-resistant virus mutant (AD169-GFP314) are shown in Fig. 2a and 2b, respectively.

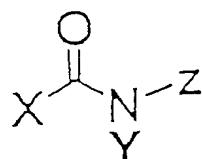
- 20 -

Claims

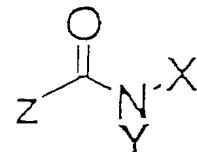
1. Use of compounds of the general formulae (Ia), (Ib), (Ic) or (Id):

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(Ia)

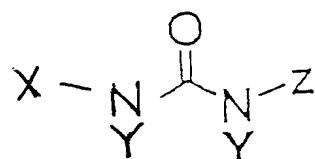


(Ib)



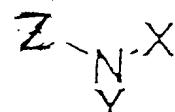
(Ic)

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(Id)

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wherein X and Z are substituents comprising an aromatic or heteroaromatic ring system and Y is hydrogen or C₁-C₄ alkyl as well as the pharmacologically acceptable salts thereof, for the preparation of an agent against infectious diseases.

- 25
2. Use of claim 1,

wherein X and/or Z is an aromatic radical which is unsubstituted or which carries at least one substituent which may be selected from hydroxy, cyano, nitro, halo, C₁-C₄ alkyl, C₁-C₄ haloalkyl, C₁-C₄ alkoxy, C₁-C₄ haloalkoxy, C₁-C₄ alkylthio, C₁-C₄ haloalkylthio, carboxy, carboxy-C₁-C₄-alkyl, carboxy-aryl or -heteroaryl,

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- 21 -

aminocarbonyl - C₁-C₄-alkyl, amino carbonyl - aryl or - heteroaryl, aryl and heteroaryl.

3. Use according to claim 1 or 2,

5 wherein X and/or Z is a phenyl radical which may carry up to 4 substituents.

4. Use according to claim 1 or 2,

wherein X and/or Z carries at least one C₁-C₃ haloalkyl substituent.

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5. Use according to claim 4,

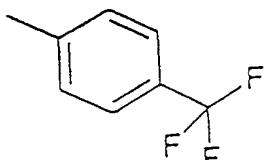
wherein X and/or Z carries at least one substituent selected from -CF₃, -CHF₂ and -CH₂F.

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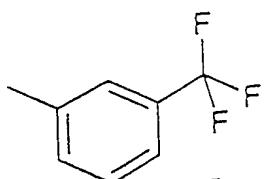
6. Use according to claim 5,

wherein X is selected from radicals represented by the formulae (IIa-c):

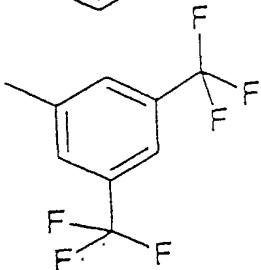
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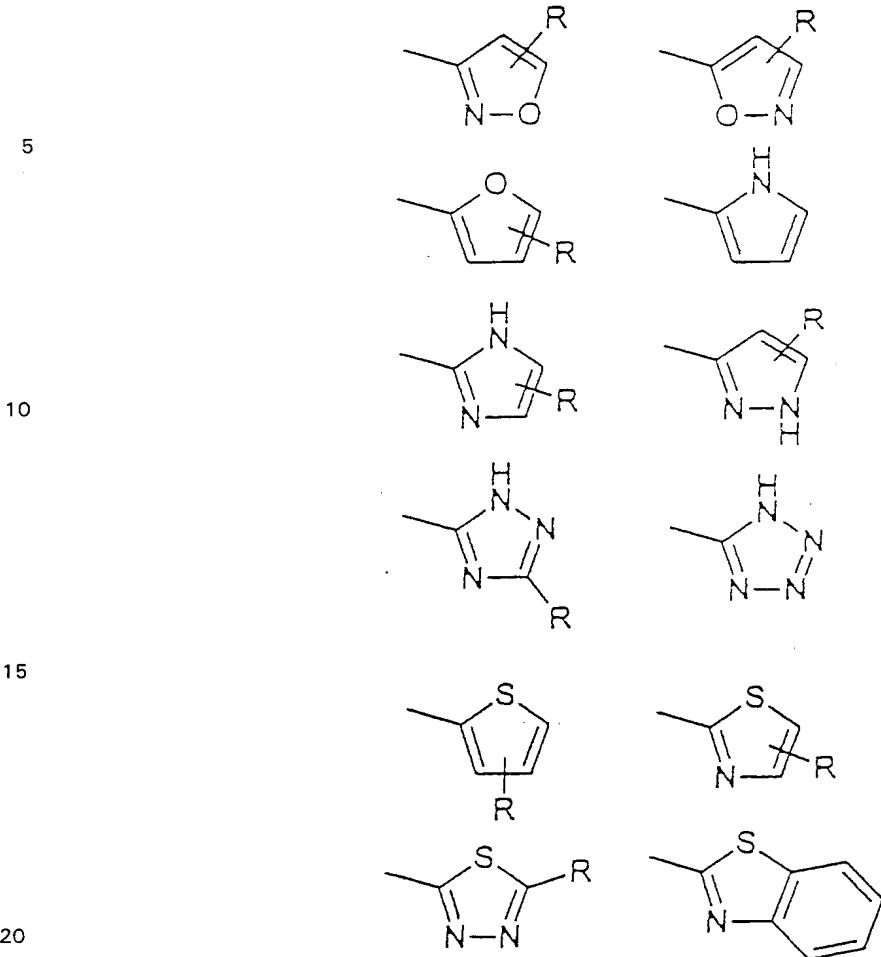
- 22 -

7. Use according to any of the claims 1 - 6,
wherein X and/or Z comprises a heterocyclic ring.
8. Use according to any one of claims 1-5,
5 wherein X and/or Z comprises a five-membered heterocyclic ring, a
six-membered heterocyclic ring or a bi- or polyheterocyclic ring.
9. Use according to claim 8,
wherein X and/or Z comprises a five-membered heterocyclic ring
10 selected from pyrrole, pyrazole, imidazole, 1,2,3-triazole, tetrazole,
oxazole, isoxazole, thiazole, isothiazole, 1,2,4-thiadiazole, 1,3,4-thiadiazole, thiophene, furan, indole and 3-thiaindole.
10. Use according to claim 9,
15 wherein X and/or Z is selected from radicals represented by the
formulae (IIIa-k):

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wherein R is at least one substituent selected from halo, C₁-C₃ alkyl and C₁-C₃ alkoxy.

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11. Use according to any one of claims 1-10,
wherein compounds (Ia), (Ib), (Ic) and (Id) have a higher selectivity
for kinases from pathogens than for the cellular enzyme DHODH.

- 30 12. Use according to any one of claims 1-11 for the preparation of an
agent against viral infections.

- 24 -

13. Use according to claim 12 for the preparation of an agent against infections by herpesviruses.

14. Use according to claim 13,

5 wherein the virus is selected from herpes simplex viruses, varicella viruses, cytomegaloviruses, muromegaloviruses, roseoloviruses, lymphocryptoviruses and rhadinoviruses.

15. Use according to claim 14,

10 wherein the virus is human cytomegalovirus (HCMV).

16. Use according to any one of claims 12-15,

 wherein the virus is a Foscarnet- or ganciclovir-resistant virus strain.

15 17. Use according to any one of claims 1-16,

 wherein the agent is suitable for oral, parenteral, rectal, nasal and topical application.

18. Use according to any one of claims 1-17,

20 wherein the dosage of the active agent is from 0.01 mg to 100 mg per day and per kg body weight.

19. Use according to any one of claims 1-18 as a combination therapy with viral DNA polymerase inhibitors.

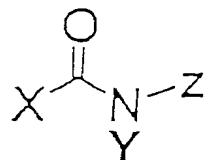
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20. A method for the treatment or prophylaxis of a herpesvirus infection comprising administering to a subject in need thereof a pharmaceutically effective amount of a compound according to general formulae (Ia), (Ib), (Ic) or (Id):

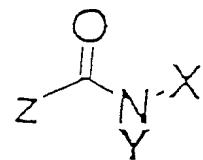
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- 25 -

5 (Ia)



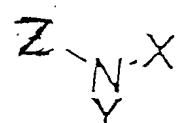
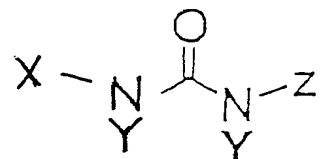
(Ib)



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(Ic)

(Id)



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wherein X and Z are substituents comprising an aromatic or heteroaromatic ring system and Y is hydrogen or C₁-C₄ alkyl.

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21. Compounds according to the general formulae (Ia), (Ib), (Ic) or (Id):

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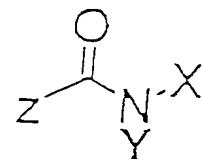
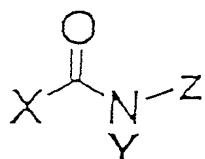
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- 26 -

(Ia)

(Ib)

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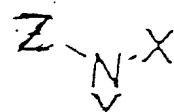
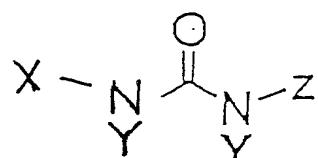


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(Ic)

(Id)

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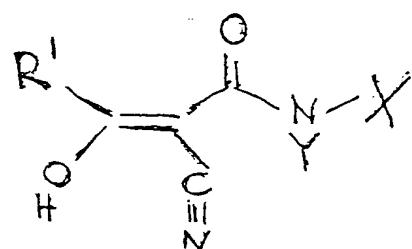


wherein X and Z are substituents comprising an aromatic or heteroaromatic ring system and Y is hydrogen or $\text{C}_1\text{-C}_4$ alkyl.

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22. Use of compounds of the general formula (IV)

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(IV)

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wherein X and Y are defined as in any one of claims 1-7 and R' is $\text{C}_1\text{-C}_4$ alkyl or $\text{C}_3\text{-C}_4$ cycloalkyl as well as the pharmacologically

- 27 -

acceptable salts thereof, for the preparation of an agent against ganciclovir-resistant virus strains.

23. Use according to claim 22,

5 wherein R' is CH₃.

24. Use according to claim 22 or 23,

wherein the virus is human cytomegalovirus (HMCV).

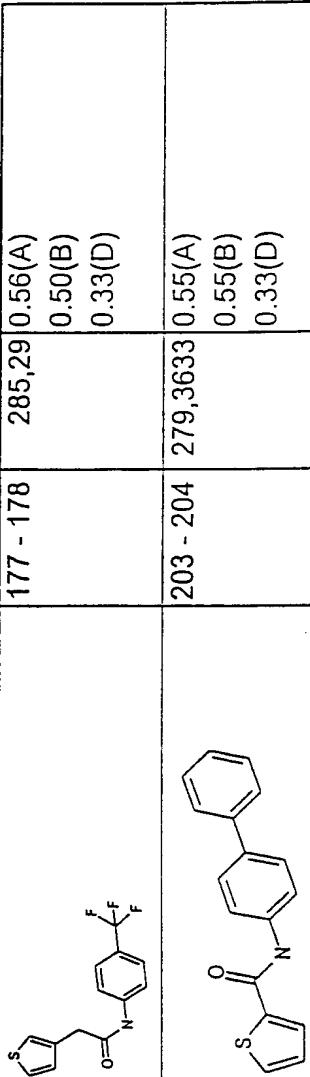
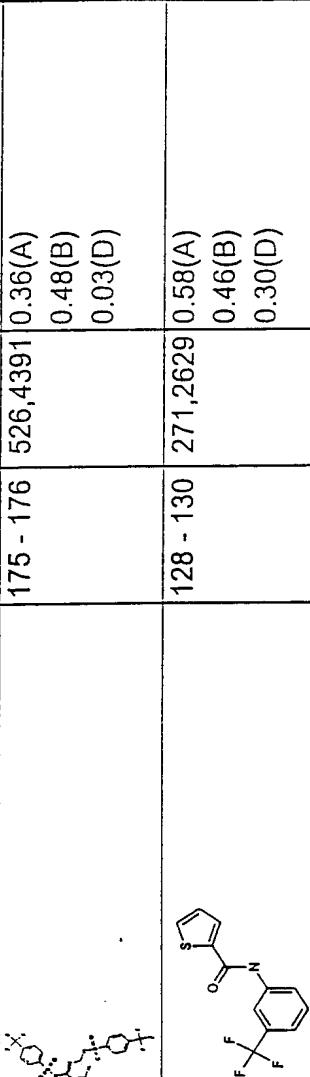
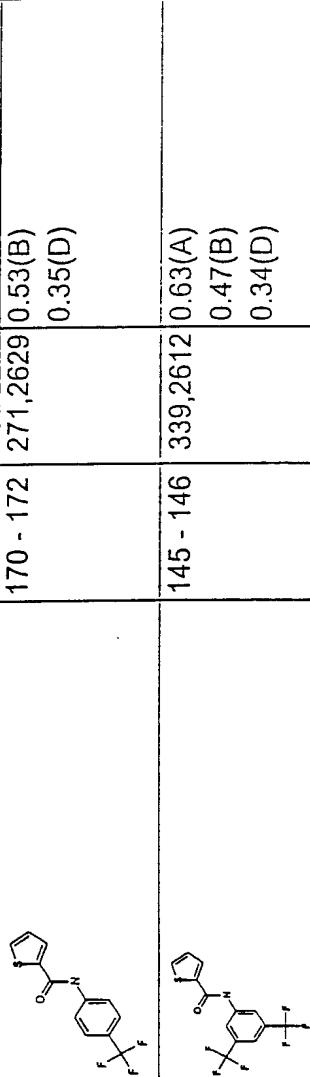
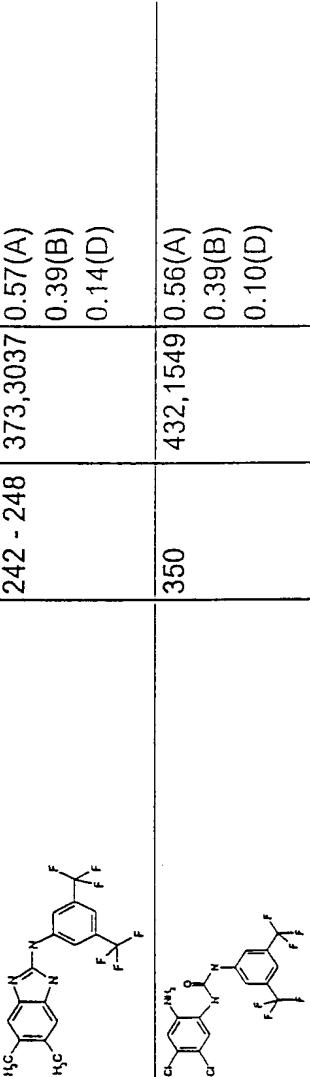
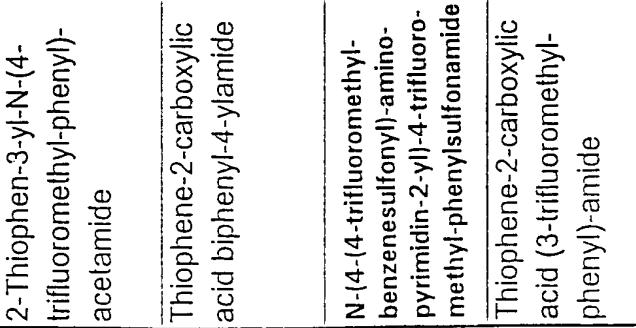
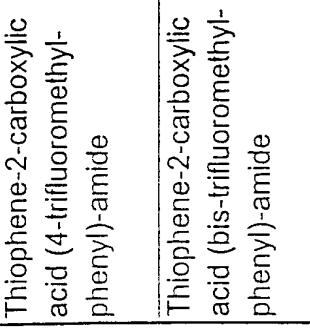
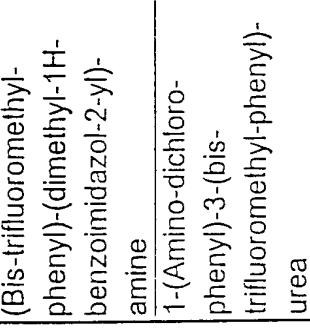
Fig. 1

IUPAC NAME	STRUCTURE	MP	mol weigh	TLC Rf.
N-(5-Phenyl-1H-pyrazol-3-yl)-4-trifluoromethylbenzamide		234 - 235	331,2999	0.59(B) 0.63(C)
4-Trifluoromethyl-N-(4-trifluoromethyl-phenyl)benzamide		104 - 106	333,2355	0.79 (B) 0.51(C) 0.78(D)
N-Quinolin-8-yl-trifluoromethylbenzamide		75 - 78	316,2852	0.93(B) 0.51(C) 0.78(D)
N-(4-Iodo-phenyl)-4-trifluoromethylbenzamide		148 - 151	391,1335	0.82(B) 0.49(C) 0.84(D)
N-(3-(4-trifluoromethyl-benzoyl)-amino-phenyl)-4-trifluoromethylbenzamide		187 - 189	452,3595	0.61(B) 0.53(D)
N-(4-Cyano-phenyl)-4-trifluoromethylbenzamide		179 - 180	290,247	0.64(B) 0.51(D)
3,4-Dimethyl-thieno[2,3-b]thiophene-2-carboxylic acid[(bis-trifluoro methyl)-phenyl]amide		118 - 120	423,4017	Maybridge KM 03523
Quinoline-6-carboxylic acid (3-trifluoromethyl-phenyl)-amide		157 - 160	316,2852	KM 10124

continued Fig. 1

Benzo[1,2,5]oxadiazole-5-carboxylic acid >(3-trifluoromethyl-phenyl)-amide			307,234	Maybridge KM 09572
Thiophene-2-carboxylic acid[3,4-dichloro-2-(3,4-dimethyl-phenyl)amide]		180 - 183 222 - 223	419,3327	0.61(A) 0.67(B) 0.42(D)
Thiophene-2-carboxylic acid[3,4-dichloro-2-(5-phenyl-1H-pyrazol-3-ylcarbamoyl)-phenyl]-amide		254 - 255	457,3413	0.35(A) 0.50(B) 0.06(D)
N-(3-Bromo-phenyl)-2-thiophen-2-yl-acetamide		98 - 100	296,1876	0.56(A) 0.54(B) 0.36(D)
N-(3-Bromo-phenyl)-2-thiophen-3-yl-acetamide		100 - 101	296,1876	0.55(A) 0.55(B) 0.34(D)
N-(4-Phenoxy-phenyl)-2-thiophen-2-yl-acetamide		98 - 104	309,3898	0.52(A) 0.58(B) 0.32(D)
Thiophene-3-carboxylic acid (4-bromo-phenyl)-amide		194 - 196	282,1605	0.56(A) 0.51(B) 0.38(D)
N-(4-Bromo-phenyl)-2-thiophen-3-yl-acetamide		150	296,1876	0.54(A) 0.51(B) 0.29(D)

continued Fig. 1

		177 - 178	285,29	0.56(A) 0.50(B) 0.33(D)
	203 - 204	279,3633	0.55(A) 0.55(B) 0.33(D)	
	175 - 176	526,4391	0.36(A) 0.48(B) 0.03(D)	
	128 - 130	271,2629	0.58(A) 0.46(B) 0.30(D)	
	170 - 172	271,2629	0.53(B) 0.35(D)	
	145 - 146	339,2612	0.63(A) 0.47(B) 0.34(D)	
	242 - 248	373,3037	0.57(A) 0.39(B) 0.14(D)	
	350	432,1549	0.56(A) 0.39(B) 0.10(D)	

continued Fig. 1

(3,5-Bis(trifluoromethyl)-phenyl)-(4-methyl-1H-benzimidazol-2-yl)-amine		232 - 235	359,2766 0.56(A) 0.40(B) 0.13(D)
Thiophene-2-carboxylic acid [2-(benzothiazol-2-ylcarbamoyl)-3,4-dichloro-phenyl]-amide		274 - 275	448,3524 0.52(A) 0.65(B) 0.18(D)
Thiophene-2-carboxylic acid [5-chloro-2-(3,4-dimethyl-phenylcarbamoyl)-phenyl]-amide		196 - 197	384,8877 0.61(A) 0.60(B) 0.31(D)
Thiophene-2-carboxylic acid [2-(4-bromo-phenylcarbamoyl)-5-chloro-phenyl]-amide		236 - 237	435,7295 0.61(A) 0.59(B) 0.29(D)
Thiophene-2-carboxylic acid [2-(4-acetylaminophenylcarbamoyl)-4-bromo-phenyl]-amide		281 - 288	458,3368 0.35(A) 0.44(B) 0.02(D)
Thiophene-2-carboxylic acid [2-(benzothiazol-2-ylcarbamoyl)-4-bromo-phenyl]-amide		252 - 255	458,3584 0.56(A) 0.65(B) 0.21(D)
Thiophene-2-carboxylic acid [2-(4-bromo-phenylcarbamoyl)-4-chloro-phenyl]-amide		218 - 220	435,7295 0.61(A) 0.63(B) 0.34(D)
Thiophene-2-carboxylic acid [2-(benzothiazol-2-ylcarbamoyl)-4-chloro-phenyl]-amide		257 - 260	413,9074 0.57(A) 0.65(B) 0.25(D)

continued Fig. 1

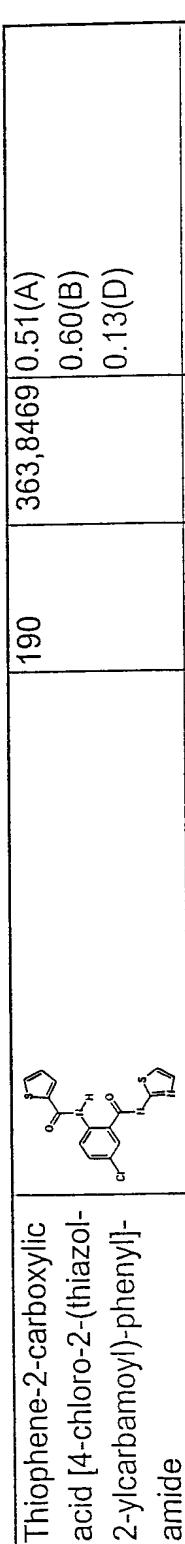
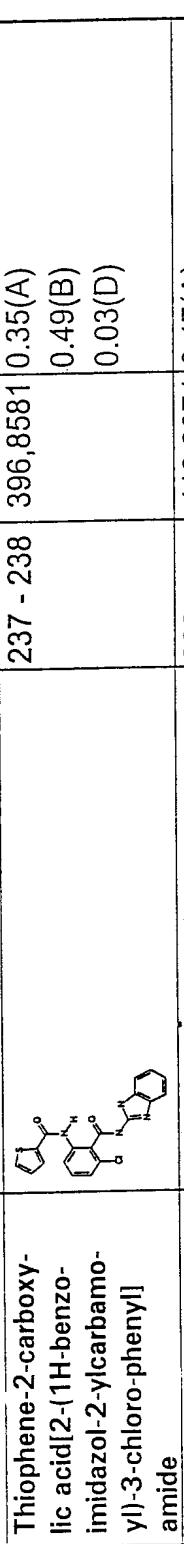
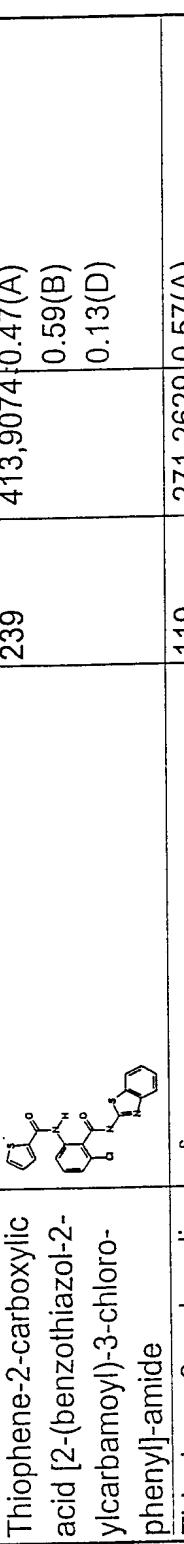
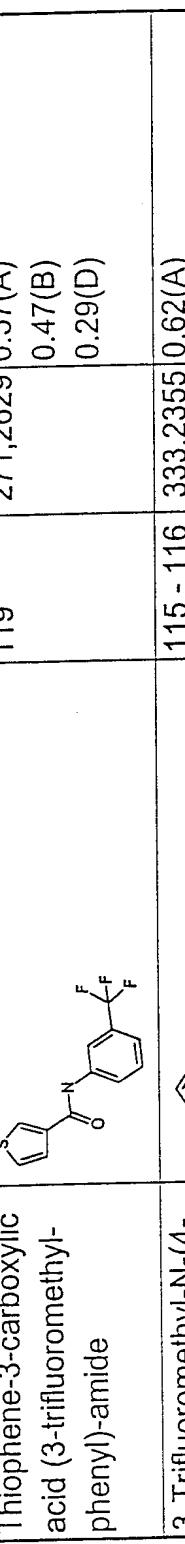
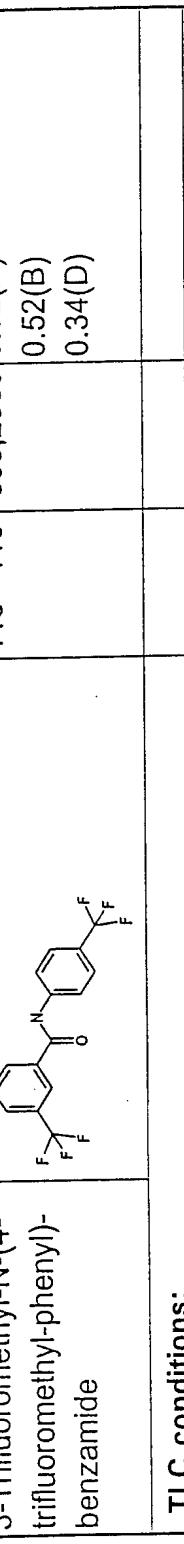
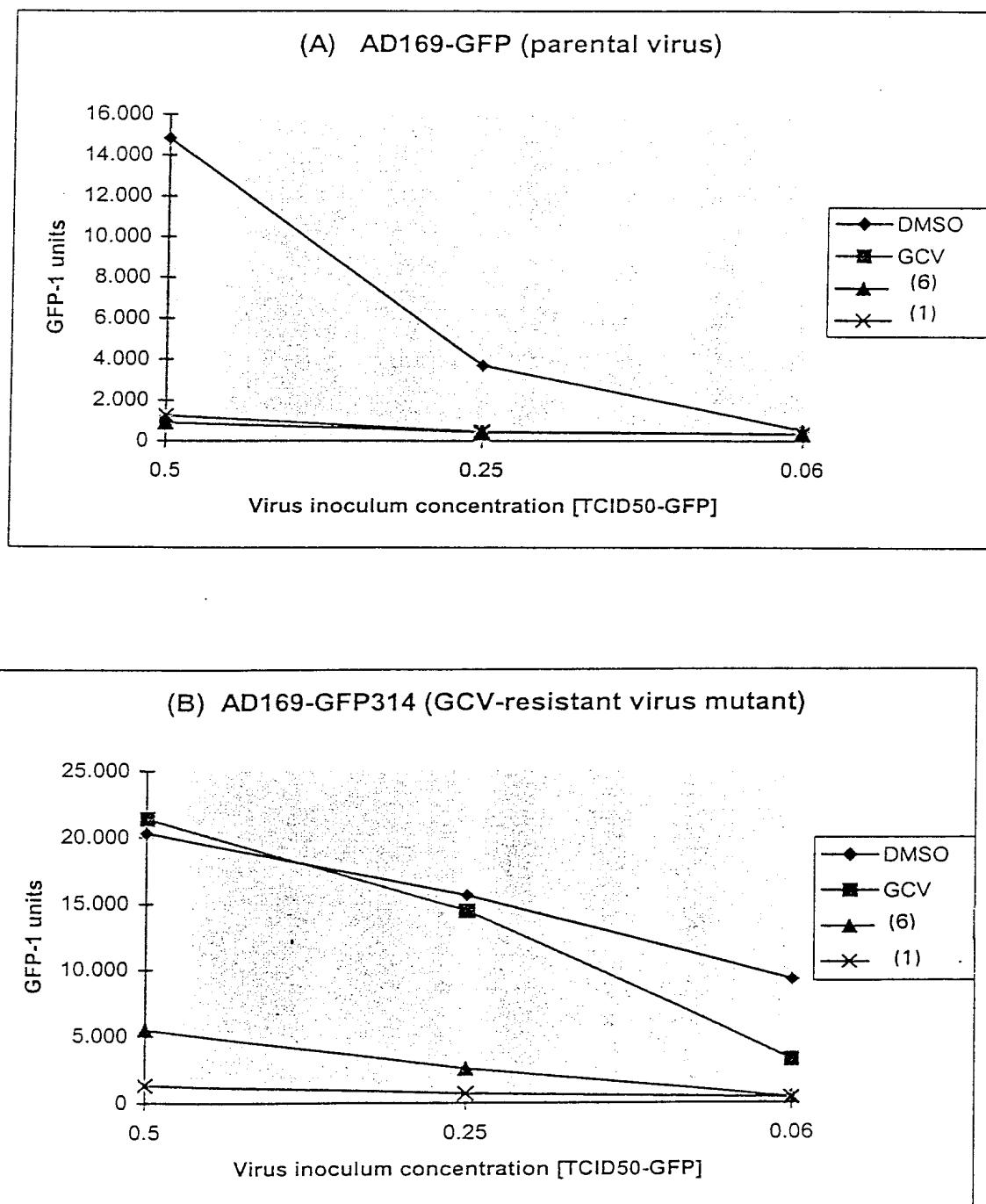
		190	363,8469	0.51(A) 0.60(B) 0.13(D)
		237 - 238	396,8581	0.35(A) 0.49(B) 0.03(D)
		239	413,9074	0.47(A) 0.59(B) 0.13(D)
		119	271,2629	0.57(A) 0.47(B) 0.29(D)
		115 - 116	333,2355	0.62(A) 0.52(B) 0.34(D)
TLC conditions:				
A: Hexane: Acetone 1:1				
B: Benzene: Methanol 1:4				
C: Ethylacetate: Hexane 1:5				
D: Ethylacetate: Hexane 1:2				

Fig. 2



(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
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(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

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31 January 2002

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 01/21160 A3

(54) Title: CARBOXYMIDE AND ANILINE DERIVATIVES AS SELECTIVE INHIBITORS OF PATHOGENS

(57) Abstract: The present invention relates to the use of carboxamide compounds as selective inhibitors of pathogens, particularly viruses and, more particularly, herpesviridae. Surprisingly, these compounds show reduced side effects in comparison with previous antiviral compounds. Thus, a novel method for preventing or treating infections by pathogens, particularly herpesviridae is provided.

INTERNATIONAL SEARCH REPORT

International Application No

PC EP 00/09306

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 7 A61K31/47 A61P31/12 A61P31/14

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHEDMinimum documentation searched (classification system followed by classification symbols)
 IPC 7 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, CHEM ABS Data, MEDLINE, EMBASE, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 40381 A (ARMISTEAD DAVID M ;FRANK CATHARINE A (US); NOVAK PERRY M (US); VER) 17 September 1998 (1998-09-17) abstract page 2, line 8 - line 9 page 6, line 10 -page 7, line 6 page 10, line 3 - line 5 table I claims 1-12,16 ---	1-21
X	EP 0 821 952 A (HOECHST AG) 4 February 1998 (1998-02-04) abstract page 1, line 24 -page 4, line 32 claims 1-10 --- -/-	1-21

 Further documents are listed in the continuation of box C. Patent family members are listed in annex.

° Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority, claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

25 January 2001

Date of mailing of the international search report

17.05.01

Name and mailing address of the ISA

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Taylor, G

INTERNATIONAL SEARCH REPORT

International Application No PC , EP 00/09306
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 96 16675 A (REGA INST) 6 June 1996 (1996-06-06) cited in the application abstract page 3, line 1 -page 5, line 1 claims 1-36 ---	1-21
X	WO 95 19169 A (MAX PLANCK GESELLSCHAFT ;BIOSIGNAL LTD (HU); YISSUM RES DEV CO (IL) 20 July 1995 (1995-07-20) cited in the application abstract page 5, line 6 -page 8, line 8 page 15, line 1 -page 24, line 9; figures 1A-1K,2A-2J ---	1-21
X	US 3 303 201 A (STECKER, HERBERT C.) 7 February 1967 (1967-02-07) cited in the application column 1, line 13 - line 26 table I claims 1-3 ---	21
P,X	WO 00 40242 A (AXXIMA PHARMACEUTICALS AG ;ULLRICH AXEL (DE)) 13 July 2000 (2000-07-13) abstract page 1, line 7 - line 12 page 3, line 4 -page 4, line 17 claims 1-13 -----	1-21

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP 00/09306

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
Although claim 20 is directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. Claims Nos.: **11** because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1 - 21 (part)

Remark on Protest

- The additional search fees were accompanied by the applicant's protest.
 No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 11

Present claim

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-21 (part)

The use of amide derivatives of the general formulae (Ia) or (Ib) (which are equivalent) for the preparation of a medicament for the treatment of infectious diseases.

2. Claims: 1-21 (part)

The use of urea derivatives of the general formula (Ic) for the preparation of a medicament for the treatment of infectious diseases.

3. Claims: 1-21 (part)

The use of amine derivatives of the general formula (Id) for the preparation of a medicament for the treatment of infectious diseases.

4. Claims: 22-24

The use of crotonamide derivatives of the general formula (IV) for the preparation of a medicament for the treatment of ganciclovir-resistant virus strains.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PC., EP 00/09306

Patent document cited in search report	Publication date	Patent family member(s)			Publication date
WO 9840381	A 17-09-1998	US 5932600 A			03-08-1999
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		US 5783592 A			21-07-1998
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(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
29 March 2001 (29.03.2001)

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(71) Applicant (for all designated States except US): **AXXIMA PHARMACEUTICALS AKTIENGESELLSCHAFT** [DE/DE]; Am Klopferspitz 19, 82152 Martinsried (DE).

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(72) Inventors; and

(75) Inventors/Applicants (for US only): **ULLRICH, Axel** [DE/DE]; Türkenstrasse 104, 80799 München (DE). **MARSCHALL, Manfred** [DE/DE]; Fuchsweg 52a, 85598 Baldham (DE). **STAMMINGER, Thomas** [DE/DE]; Gleiwitzerstrasse 5, 90522 Oberasbach (DE). **WALLASCH, Christian** [DE/DE]; Stiftsbogen 130, 81375 München (DE). **OBERT, Sabine** [DE/DE]; Bellinzonastrasse 17/2, 81475 München (DE).



WO 01/21160 A3

(54) Title: CARBOXYMIDE AND ANILINE DERIVATIVES AS SELECTIVE INHIBITORS OF PATHOGENS

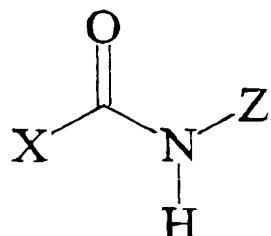
(57) Abstract: The present invention relates to the use of carboxamide compounds as selective inhibitors of pathogens, particularly viruses and, more particularly, herpesviridae. Surprisingly, these compounds show reduced side effects in comparison with previous antiviral compounds. Thus, a novel method for preventing or treating infections by pathogens, particularly herpesviridae is provided.

AMENDED CLAIMS

[received by the International Bureau on 17 July 2001 (17.07.01);
original claims 1-24 replaced by amended claims 1-16 (5 pages)]

1. Use of a compound of the general formula (I):

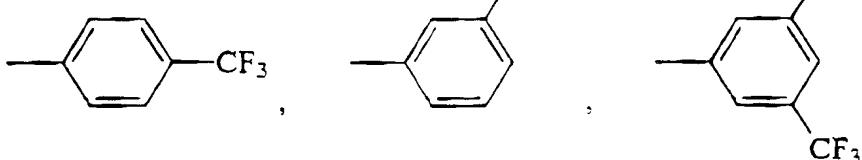
5



10

wherein

X represents



15

or a substituent comprising a thiophene ring system;

20 Z is a substituent comprising an aromatic ring system, or a six-membered heterocyclic ring, or a bicyclic heterocyclic ring, or a five-membered heterocyclic ring wherein said five-membered heterocyclic ring is selected from the group comprising pyrrole, pyrazole, imidazol, oxazole, isoxazole, thiazole, isothiazole, thiophene, furan, indole and 3-thiaindole;

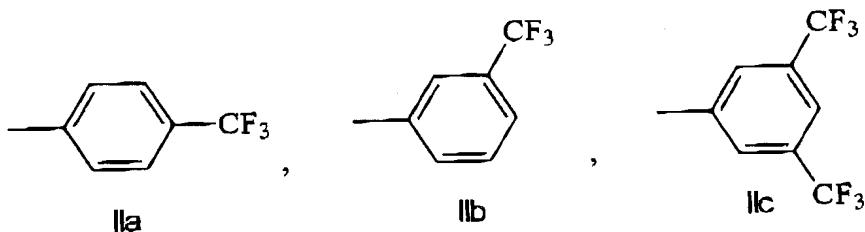
25 under the proviso that Z does not represent a phenyl ring substituted with up to three further substituents selected from the group consisting of -Cl, -Br, -I, and -CF₃, when X represents a thiophene-2-yl residue;

30 as well as pharmacologically acceptable salts thereof, for the preparation of an agent against infections by herpesviruses.

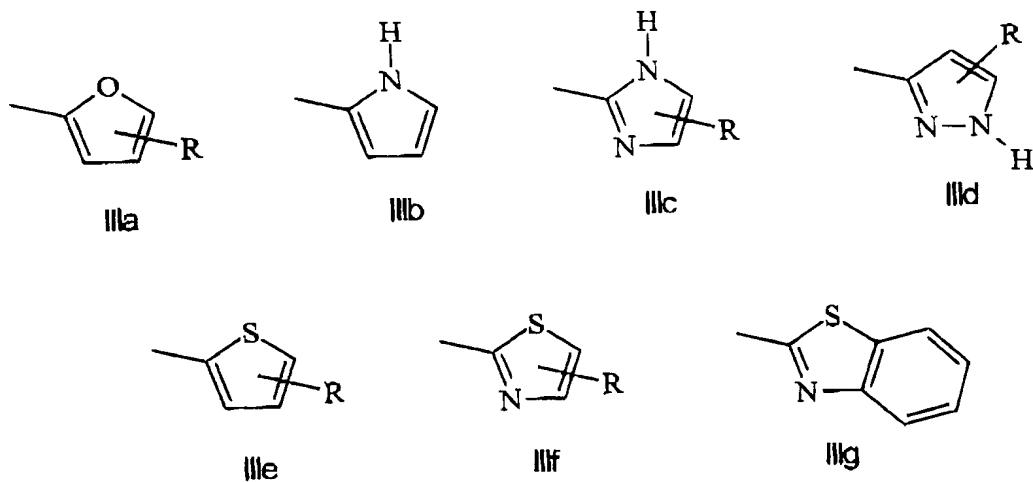
2. Use of claim 1,

35 wherein Z is an aromatic radical which is unsubstituted or which carries at least one substituent which may be selected from hydroxy, cyano, nitro, halo, C₁ – C₄ alkyl, C₁ – C₄ haloalkyl, C₁ – C₄ alkoxy, C₁ – C₄ haloalkoxy, C₁ – C₄ alkylthio, C₁ – C₄ haloalkylthio, carboxy, carboxy C₁ – C₄ alkyl, carboxyaryl, carboxyheteroaryl, aminocarbonyl C₁ – C₄ alkyl, aminocarbonylaryl, aminocarbonylheteroaryl, aryl, and heteroaryl.

3. Use according to claim 1 or 2,
wherein Z is a phenyl radical which may carry up to 4 substituents.
- 5 4. Use according to claim 1 or 2,
wherein Z carries at least one C₁ – C₃ haloalkyl substituent.
- 10 5. Use according to claim 4,
wherein Z carries at least one substituent selected from –CF₃, –CHF₂, and
–CH₂F.
- 15 6. Use according to claim 5,
wherein Z is selected from radicals represented by the formulae (IIa-c):



- 20 7. Use according to claim 1,
wherein Z is selected from radicals represented by the formulae (IIIa-j):



35 wherein R is at least one substituent selected from halo, C₁ – C₃ alkyl, and
C₁ – C₃ alkoxy.

8. Use of a compound of claim 1 wherein the compound is selected from the group comprising:
- N-(5-Phenyl-1H-pyrazol-3-yl)-4-trifluoromethyl-benzamide,
4-Trifluoromethyl-N-(4-trifluoromethyl-phenyl)-benzamide,
5 N-Quinolin-8-yl-trifluoromethyl-benzamide,
N-(4-Iodo-phenyl)-4-trifluoromethyl-benzamide,
N-(3-(4-trifluoromethyl-benzoyl)-amino-phenyl)-4-trifluoromethyl-benzamide,
N-(4-Cyano-phenyl)-4-trifluoromethyl-benzamide,
10 3,4-Dimethyl-thieno[2,3-b]thiophene-2-carboxylic acid [(bis-trifluoro-methyl)-phenyl]-amide,
Quinoline-6-carboxylic acid (3-trifluoromethyl-phenyl)-amide,
Benzo[1,2,5]oxadi-azole-5-carboxylic acid [(3-trifluoromethyl)-phenyl]-amide,
Thiophene-2-carboxylic acid [3,4-dichloro-2-(3,4-dimethyl-phenylcarbamoyl)-phenyl]-amide,
15 Thiophene-2-carboxylic acid [3,4-dichloro-2-(5-phenyl-1H-pyrazol-3-ylcarbamoyl)-phenyl]-amide,
N-(3-Bromo-phenyl)-2-thiophen-2-yl-acetamide,
N-(3-Bromo-phenyl)-2-thiophen-3-yl-acetamide,
20 N-(4-Phenoxy-phenyl)-2-thiophen-2-yl-acetamide,
Thiophene-3-carboxylic acid (4-bromo-phenyl)-amide,
N-(4-Bromo-phenyl)-2-thiophen-3-yl-acetamide,
2-Thiophen-3-yl-N-(4-trifluoromethyl-phenyl)-acetamide,
Thiophene-2-carboxylic acid biphenyl-4-ylamide,
25 (Bis-trifluoromethyl-phenyl)-(dimethyl-1H-benzoimidazol-2-yl)-amine,
Thiophene-2-carboxylic acid [2-(benzothiazol-2-ylcarbamoyl)-3,4-dichloro-phenyl]-amide,
Thiophene-2-carboxylic acid [5-chloro-2-(3,4-dimethyl-phenylcarbamoyl)-phenyl]-amide,
30 Thiophene-2-carboxylic acid [2-(4-bromo-phenylcarbamoyl)-5-chloro-phenyl]-amide,
Thiophene-2-carboxylic acid [2-(4-acetylamo-phenylcarbamoyl)-4-bromo-phenyl]-amide,
Thiophene-2-carboxylic acid [2-(benzothiazol-2-ylcarbamoyl)-4-bromo-phenyl]-amide,
35 Thiophene-2-carboxylic acid [2-(4-bromo-phenylcarbamoyl)-4-chloro-phenyl]-amide,

Thiophene-2-carboxylic acid [2-(benzothiazol-2-ylcarbamoyl)-4-chlorophenyl]-amide.

Thiophene-2-carboxylic acid [4-chloro-2-(thiazol-2-ylcarbamoyl)-phenyl]-amide.

5 Thiophene-2-carboxylic acid [2-(1H-benzoimidazol-2-ylcarbamoyl)-3-chloro-phenyl]-amide,

Thiophene-2-carboxylic acid [2-(benzothiazol-2-ylcarbamoyl)-3-chloro-phenyl]-amide,

10 Thiophene-3-carboxylic acid (3-trifluoromethyl-phenyl)-amide, and
3-Trifluoromethyl-N-(4-trifluoromethyl-phenyl)-benzamide.

9. Use according to any one of claims 1 – 8,
wherein compounds (I) have a higher selectivity for kinases from pathogens
than for the cellular enzyme DHODH.

10. Use according to claim 1,
wherein the virus is selected from herpes simplex viruses, varicella viruses,
cytomegaloviruses, muromegaloviruses, roseoloviruses,
lymphocryptoviruses and rhadinoviruses.

11. Use according to claim 10,
wherein the virus is human cytomegalovirus (HCMV).

12. Use according to any one of claims 1, 10, or 11,
wherein the virus is a Foscarnet- or ganciclovir-resistant virus strain.

13. Use according to claim 1,
wherein the agent is suitable for oral, parenteral, rectal, nasal and topical application.

14. Use according to claim 1 or 13,
wherein the dosage of the active agent is from 0.01 mg to 100 mg per day
and per kg body weight.

- 35 15. Use according to any one of claims 1 – 14,
as a combination therapy with viral DNA polymerase inhibitors.

16. Method for the treatment or prophylaxis of a herpesvirus infection comprising administering to a subject in need thereof a pharmaceutically effective amount of a compound according to general formula (I).